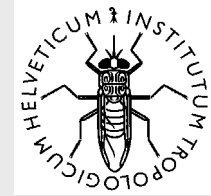


# Methods in Parasitology

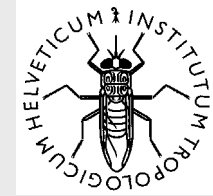
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## 5. Fresh Stool Examination (Wet mount)

Swiss Tropical Institute, Basel

April 2005



## 5.0 Fresh Stool Examination

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- Contents
- 5.1 Materials
- 5.2 Preparing wet mount
- 5.7 Examining wet mount

Production Team:

Yvette Endriss, Elisabeth Escher & Birgit Rohr  
Prof. Hanspeter Rohr (NeoCortex Foundation)  
Prof. Niklaus Weiss (STI)

## 5.1 Fresh Stool Examination

### Materials:

- Microscope slides
- Cover slips
- Sodium chloride solution in small bottle with pipette
- Wooden stick
- Fresh stool
- Gloves



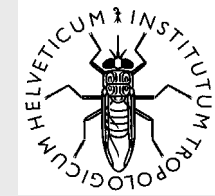
## 5.2 Fresh Stool Examination



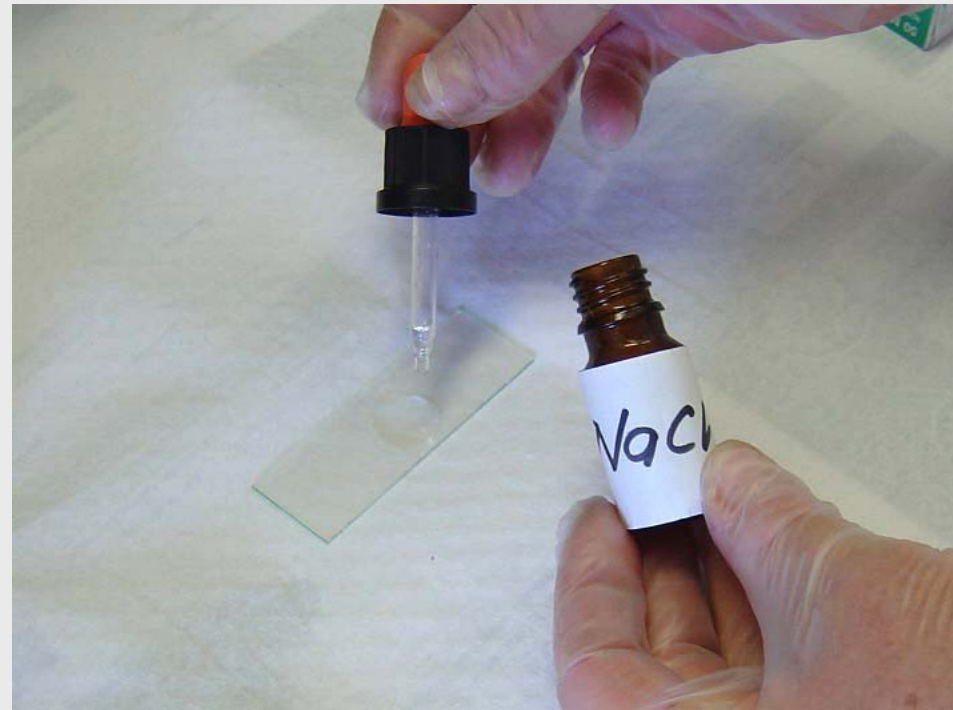
Use cleaned microscope slides



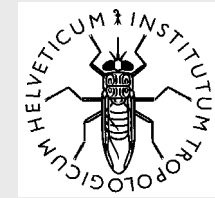
## 5.3 Fresh Stool Examination



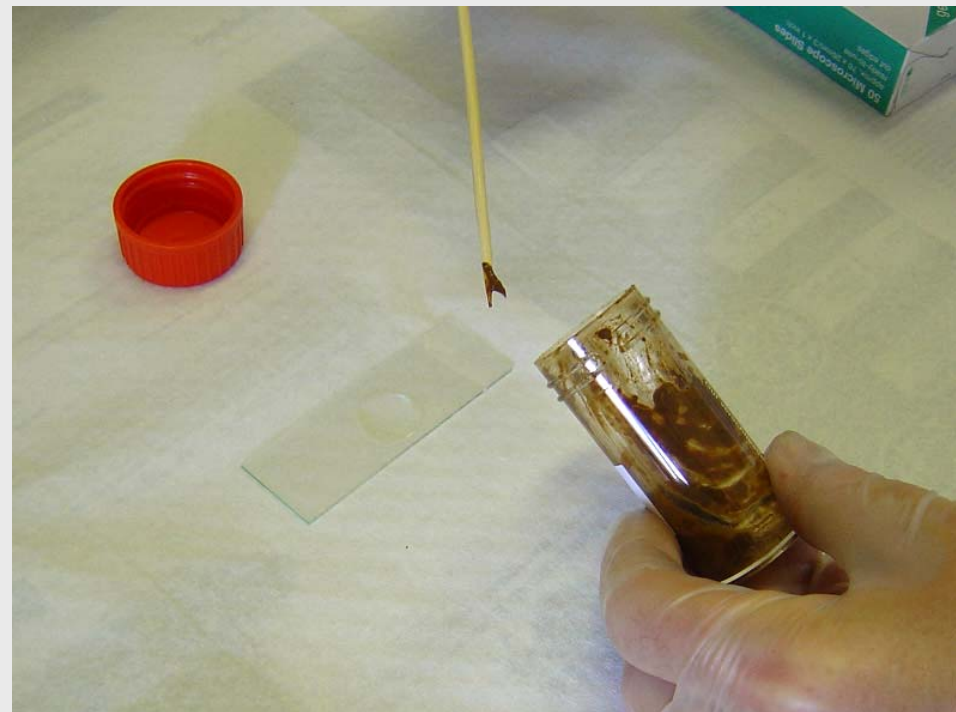
Place a drop of saline



## 5.4 Fresh Stool Examination



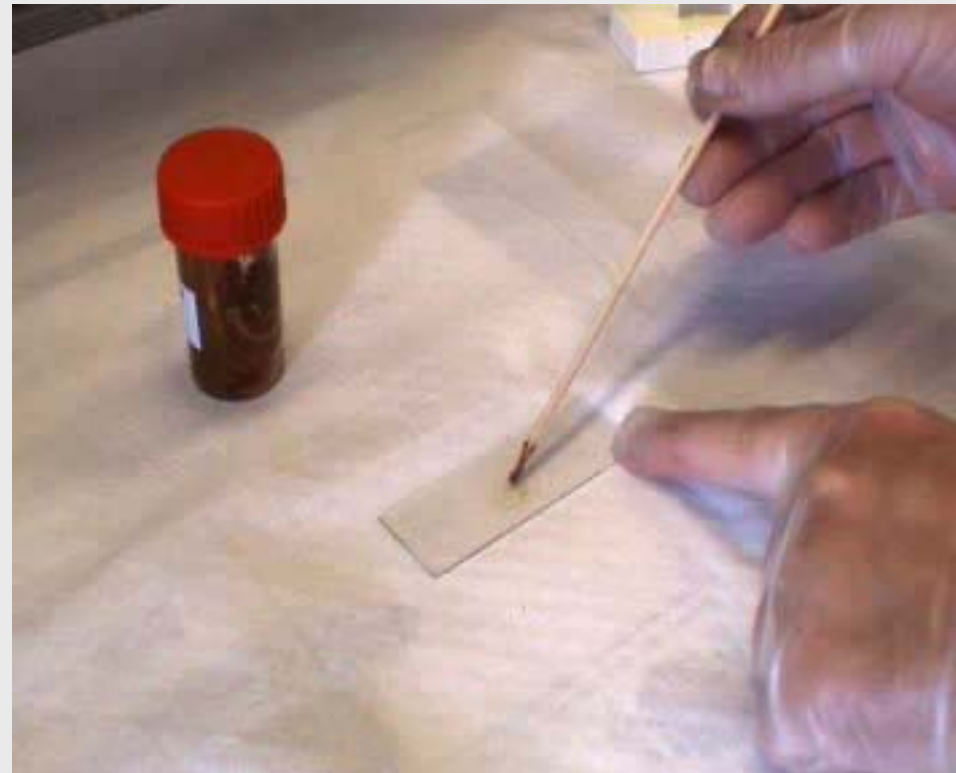
Take a small amount of stool with a wooden stick



## 5.5 Fresh Stool Examination



Mix stool with saline

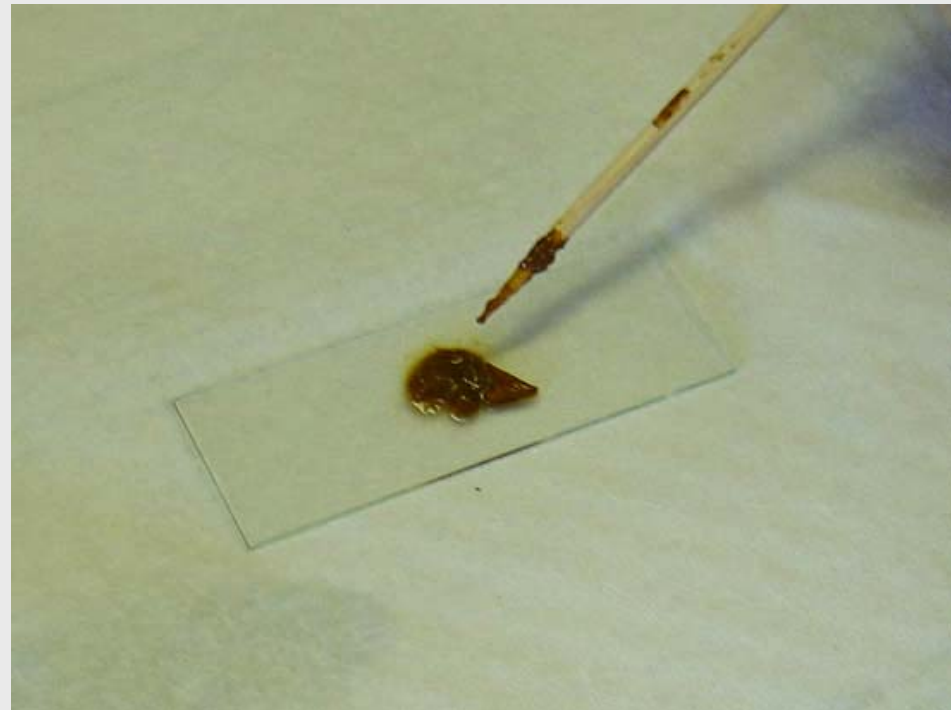


Clip

## 5.6 Fresh Stool Examination



Mistake:  
Too much stool!

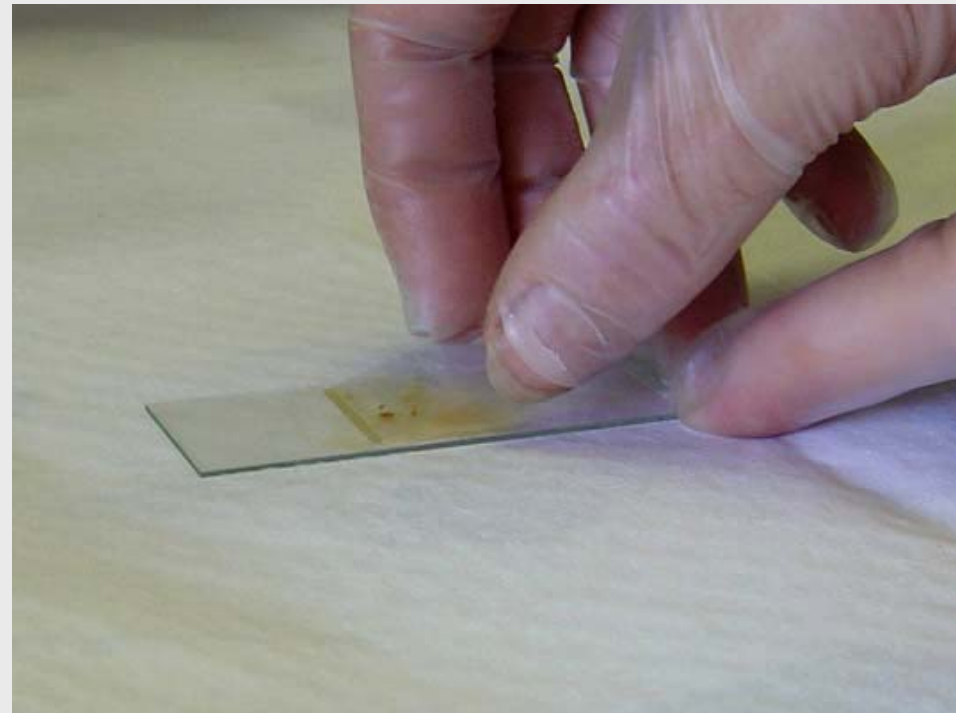


## 5.7 Fresh Stool Examination

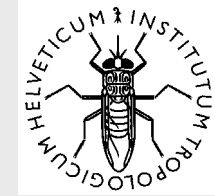


Place coverslip

Avoid air bubbles!



## 5.8 Fresh Stool Examination



Examination of helminth  
ova :  
Use 10x objective

Photo:  
*Schistosoma mansoni*





## 5.9 Fresh Stool Examination

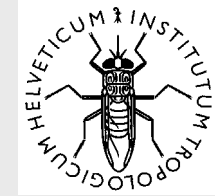
Examination of amoebae:

Press cover slip slightly, remove excess liquid with paper towel



Clip

## 5.10 Fresh Stool Examination



For amoebae:

Use **50x** objective with  
oil immersion

Example:

*Entamoeba coli*, cyst

